



## To study the effect of cadmium chloride on the level of oxidative stress markers in *Channa punctatus*

Dr. C Manjusha<sup>1</sup>, Hajera Fatima<sup>2</sup>, Dr. M Nageshwar<sup>3</sup>, Madhumathi Kondoor<sup>1</sup>

<sup>1</sup> Lecturer, Department of Zoology, Sarojini Naidu Vanita Maha Vidyalaya, Telangana, India

<sup>2</sup> Student, Sarojini Naidu Vanita Maha Vidyalaya, Telangana, India

<sup>3</sup> Lecturer, Department of Zoology, Telangana Mahila Viswavidyalayam, Telangana, India

### Abstract

To know the effects of cadmium chloride concentrations on the level of Lipid peroxidation (LPO) and Superoxide dismutase (SOD) in Liver and Brain tissues of fresh water murrel, *Channa punctatus*.

The fishes were exposed to cadmium chloride with two different concentrations (20ppm and 40ppm) and the oxidative stress was assessed on 1<sup>st</sup> day (24hrs), 5<sup>th</sup> day & 10<sup>th</sup> day of exposure along with the control group of fishes.

LPO elevates significantly in liver tissue on 1<sup>st</sup>, 5<sup>th</sup> & 10<sup>th</sup> day followed by brain tissue, when compared to control group.

An increased LPO level is due to formation of more lipid radicals.

Superoxide dismutase (SOD) activity decreases initially on 1<sup>st</sup> day of exposure of fishes to cadmium chloride when compared with control group of fishes and later it increases in experimental group on 5<sup>th</sup> day & 10<sup>th</sup> day in liver tissue followed by brain tissue.

Thus, fluctuation in antioxidant enzyme SOD was seen.

The results indicate that cadmium chloride may induce toxic effects which can be assessed by the oxidative stress biomarker SOD and LPO in *Channa punctatus*.

**Keywords:** antioxidant, superoxide dismutase, oxidative stress

### Introduction

Cadmium is one of the contaminants of ecosystem.

The pollution of heavy metals increases with human activities like agricultural activities and motor vehicles are all major sources of metal pollution. Metals can be found in water and sediments they are accumulated in aquatic organisms such as fish [12].

Fishes are considered to be an important nutrition to human, but those from contaminated sites when consumed are of potential risk to the human health.

*Channa punctatus* have a more commercial significance as a food fish (Parveen *et al.*, 2021).

Murrel being typical “live fishes” and soft flesh without fat are considered to have medicinal and nutritional value.

Fishes are the bio indicators of aquatic ecosystem.

The oxidative stress induced by the metal can be assessed by the oxidative damage [11].

### Cadmium

Cadmium is one of the most toxic heavy metal found in the environment and it is extremely toxic even at low concentrations, it does not serve a beneficial biological function in higher organism [25].

Human activities like combustion of fossil fuels, agricultural land and mining waste, contribute to cadmium contamination in the environment [27].

Cadmium is used as a stabilizer of PVC, pigments, alloys and in the electroplating used to protect steel from corrosion [7, 29].

In aquatic ecosystem the presence of cadmium, is regarded as a potential hazard for the aquatic organisms.

Cadmium is readily absorbed by organisms directly from the water in its free ionic form Cd<sup>2+</sup> [2].

It also affects antioxidant enzymes especially SOD and LPO [6].

### Oxidative stress

Oxidative stress is due to the imbalance between the free radicals and the antioxidants LPO & SOD in living organisms [15].

### Free radicals

Free radicals on reacting with other molecules can form new radicals.

Free radicals are harmful in high production, during physical exercise cause muscular fatigue, aging and many diseases [3].

Increased production of free radicals can cause oxidation of lipids, proteins and alterations of gene expression [11].

### Antioxidants

An antioxidant is a substance that helps to reduce oxidative stress by forming a less reactive radical or by reducing the damage of FR chain reaction on substrates such as lipids, proteins, carbohydrates or DNA [4].

Antioxidants are of two types: Enzymatic and Non-Enzymatic [18].

**Table 1:** Types of antioxidants

Enzymatic antioxidants	Non-enzymatic antioxidants
Superoxide Dismutase (SOD)	Vitamin A
Lipid Peroxidase (LPO)	Vitamin E
	Flavonoids

### Oxidative stress & Cadmium toxicity

Cd concentration increases in living organisms including human with age and diet [21].

The absorption and accumulation of these heavy metals depends upon the factors such as sex, age, health condition and exposed people (Satarug S *et al* 2004) [16].

Cd is one of the most toxic heavy metals. Its toxicity can be considered as multidirectional. Exposure of Cd leads to an increase in the production of reactive oxygen species (ROS) namely hydrogen peroxide, superoxide radicals and hydroxyl radicals. Oxidative stress is induced by Cd [14].

The oxidative stress results in the oxidation and damage of Deoxyribonucleic acid (DNA), proteins and lipids and phospholipids of cellular membrane. Hence by lowering the mitochondrial potential Cd disrupts the synthesis of adenosine tri phosphate (ATP) and oxidative phosphorylation [28].

The ROS production is balanced by enzymatic (SOD and CAT) and non-enzymatic (Vit C, Vit E, GSH) antioxidant barriers.

Cd also damages DNA and also inhibits DNA repair enzymes [17].

It is also influenced with gene expression by disrupting cell signalling pathway. It also causes apoptosis and necrosis (Wang, Liu, *et al.*, 2013).

Cadmium has an adverse effect on the respiratory, reproductive and haematological system in many fishes [24].

### Oxidative stress biomarkers

Biomarkers serve an important role in bio monitoring of toxic metal pollution in aquatic ecosystem by indicating the oxidative stress in various organisms [23]. From a clinical point of view, the use of biomarkers to assess the degree of oxidative stress is very valuable.

### Lipid per oxidation (LPO)

Lipid per Oxidation is an antioxidant enzyme and also a biomarker of pollutants exposure in various organisms [11].

Lipid peroxidation results in the formation of lipid radicals and this results in the formation of complex mixture of lipid degradation products [1].

Heavy metal pollution in aquatic organisms is induced by LPO [5].

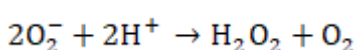
The antioxidant enzyme protects against oxidative stress and also to tissue specific damage. These antioxidant enzymes are also a biomarker of ROS and a tool in environmental assessment risk factors [8].

The ROS can be protected and detoxified by antioxidant enzymes such as lipids, proteins and DNA damage [13].

### Superoxide dismutase (SOD)

SOD is the antioxidant enzyme that can cause a catalytic reaction by dismutation of superoxide anion to O<sub>2</sub> and H<sub>2</sub>O<sub>2</sub> and CAT reacts with H<sub>2</sub>O<sub>2</sub> to form water and oxygen molecule [10, 13].

SOD can cause defence upon superoxide radicals and is the first line of defence against oxidative stress. The dismutation of O<sub>2</sub><sup>-</sup> and formation of H<sub>2</sub>O<sub>2</sub> can be catalyzed by the enzyme SOD.



### Aims & Objectives

#### Aim

To study the effect of cadmium chloride toxicity on the level of Oxidative stress markers LPO and SOD in liver and brain tissues of *Channa punctatus*.

#### Objectives

To study the effect of cadmium chloride on liver and Brain tissues of *Channa punctatus* by estimating the level of oxidative stress markers (LPO & SOD).

To study the effect of cadmium chloride on 1<sup>st</sup> day, 5<sup>th</sup> day and 10<sup>th</sup> day of exposure to fishes (*C. punctatus*) and compare the results obtained with control values.

### Experimental design

#### Materials

Tubs

Net

Fishes

Dissection box

#### Collection of fish sample

Edible fish samples of *Channa punctatus* of same size and weight will be collected from nearby Fish market, Hyderabad, Telangana.

The fishes will be collected in air filled polythene bags with water for the Experiment to be estimated in the Laboratory, for further analysis.

#### *Channa punctatus*

It is a fresh water murrel, snake-headed belonging to family Channidae of the order perciformes.

*Channa punctatus* is a fresh water murrel lives in rivers, lakes, ponds, swamps and marshes.

They feed on invertebrates and small fishes (Narejo *et al.*, 2005, 2010).

It is a bottom dweller geographically distributed important economic food source to human.

The presence of respiratory organ helps fish to survive in difficult conditions.

Murrel fingerlings are used for giving medicine every year in month of June on the eve of mrigasira karthi for allotment of asthma patients that gather all over the country in Hyderabad since long back.

It can be maintained *in vitro* condition. It is a good model for studying of biochemical effects of certain pollutants.

*Channa punctatus* has ability to respond to environmental pollution.

*Channa punctatus* was selected because of its ecotoxicological characteristics

Fresh water murrel can be easy acclimatization to laboratory conditions and availability throughout the season [9].

### Methodology

To Estimate the effect of cadmium chloride on oxidative stress biomarkers by

- Lipid peroxidation method by Garcia *et al.* (2005).
- Superoxide dismutase by Marklund and Marklund (1974).

In the present study the fishes were treated with 0.05% KMnO<sub>4</sub> solution for 20 min to avoid any dermal infections and after that the fishes were acclimatized for 1week under laboratory conditions before start of the experiment to

remove the residual effects of other chemicals (Pandey *et al.*, 2006).

Stored water was used to allow the chlorine to settle as it will also cause stress to fishes. water is changed daily to maintain same concentrations of cadmium.

Three tubs were taken and they were filled with 5 litres of water, 15 fishes were taken, they were divided in to three groups, with 5 fishes in each group. One group is taken as control, second group and third groups are taken as experimental in which fishes were exposed to cadmium chloride with two different concentrations for a time period of the 1<sup>st</sup> day, 5th day, & 10<sup>th</sup> day.

On each experiment day, fish was sacrificed and dissected for the removal of target organ and the experiments was performed to know the level of LPO and SOD in liver and brain tissues of *Channa punctatus*.

**Procedure**

**Lipid peroxidation (LPO)**

Lipid peroxidation in tissue was measured by modified method of Garcia *et al.* (2005). 1 mL of 10% homogenate was added to 1 mL of 20% TCA and heated at 70 OC for 10 min, and cooled at room temperature and centrifuged at 3000 rpm for 10 min. 400 µL of supernatant was mixed with 200 µL of 0.5% TBA reagent in a test tube covered with a glass marble and heated in a boiling water bath for 10 mins

and the tubes cooled to room temperature. The absorbance of the pink coloured trimethine condensation product was measured at 533 using a spectrophotometer. The results were expressed as nano mole MDA/gm weight of tissue.

**Superoxide dismutase (SOD)**

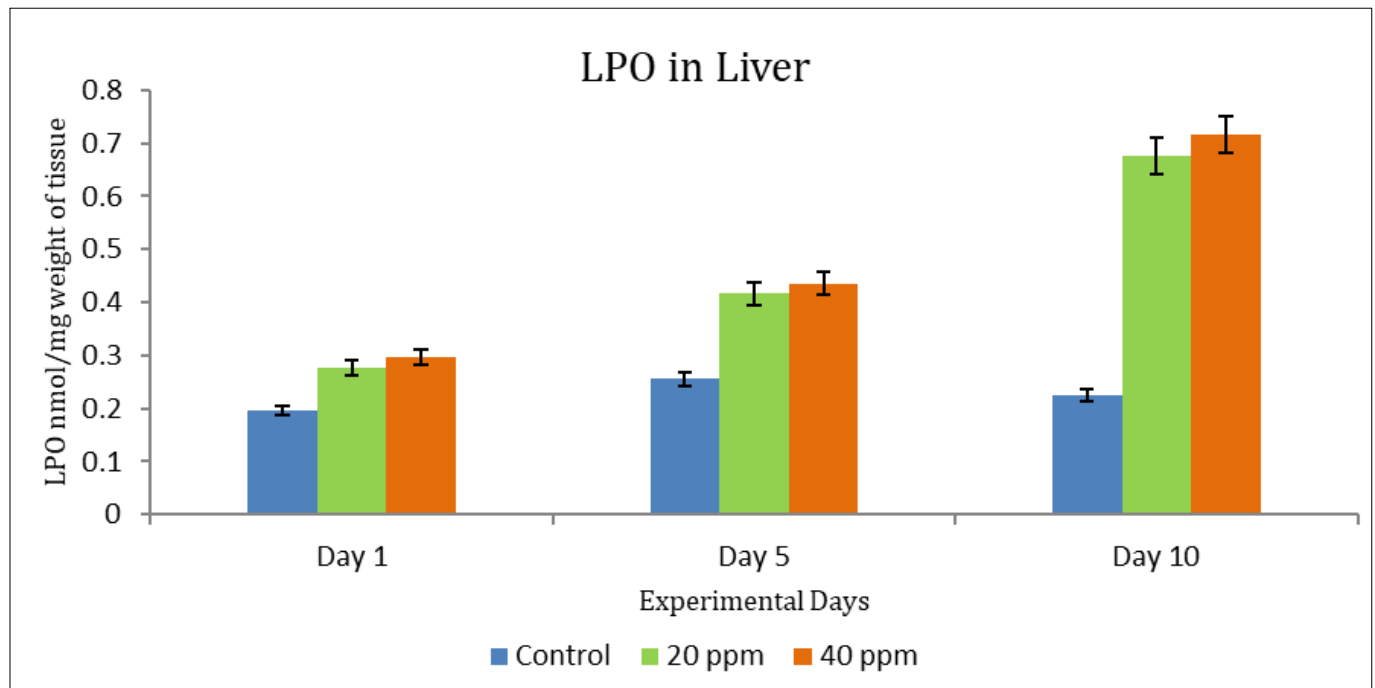
Superoxide dismutase activity in tissue was estimated by modified protocol of Marklund and Marklund (1974). The assay system in a final volume of 1.0 mL consisting 600 µL of 83.3 mM Tris-Hcl buffer, (Ph 8.2), 100 µL of 0.5 mM DETPA, 50 µL of enzyme, 50 µL of Tris-EDTA, 50 µL of 0.01 N HCl, 100 IL of H2O2 was mixed well and the reaction was initiated by adding the 50 µL of 3.97 mM Pyrogallol. Increased absorbance was read at 420 using a spectrophotometer. The enzyme activity was expressed as Units/mg protein.

**Result**

**LPO in liver**

**Table 2:** LPO level in Liver of *Channa punctatus*

Tissue	Days	Control	Experimental 20 ppm	40 ppm
Liver	1 <sup>st</sup>	0.20	0.28	0.30
	5th	0.26	0.42	0.44
	10th	0.23	0.68	0.72



**Fig 1:** Comparative chart of LPO level in liver of *Channa punctatus*

**LPO in brain**

**Table 3:** LPO level in Brain of *Channa punctatus*

Tissue	Days	Control	Experimental	
			20 ppm	40 ppm
Brain	1 <sup>st</sup>	0.20	0.22	0.25
	5th	0.26	0.36	0.38
	10th	0.23	0.44	0.47

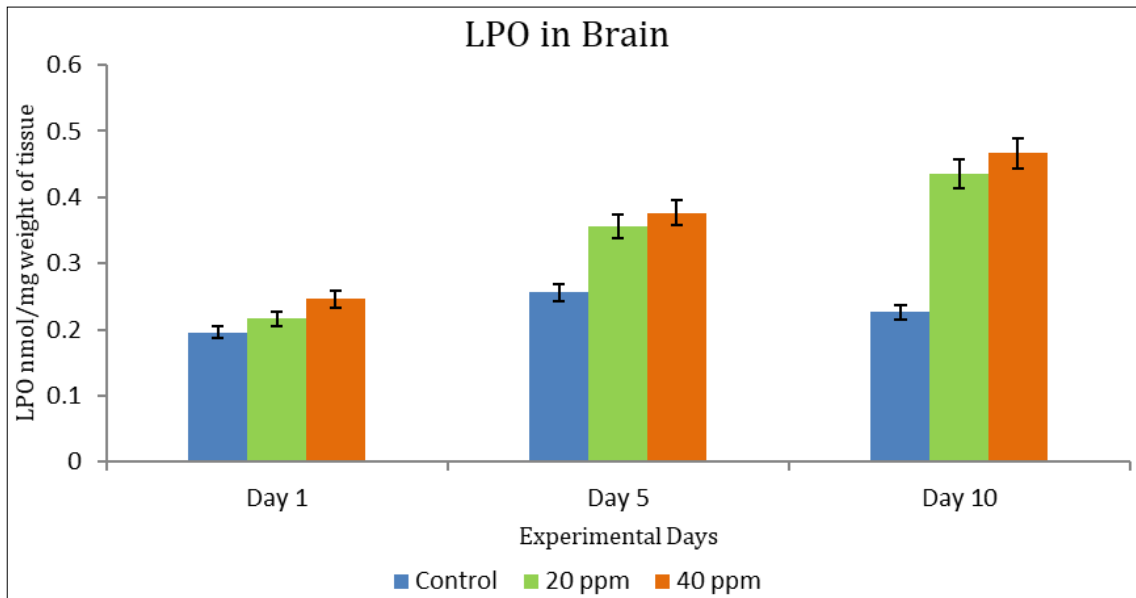


Fig 2: Comparative chart of LPO level in brain of *Channa punctatus*

**SOD in Liver**

Table 4: SOD level in liver of *Channa punctatus*

Tissue	Days	Control	Experimental	
			20 ppm	40 ppm
Liver	1 <sup>st</sup>	0.25	0.18	0.26
	5 <sup>th</sup>	0.22	0.20	0.35
	10 <sup>th</sup>	0.24	0.22	0.37

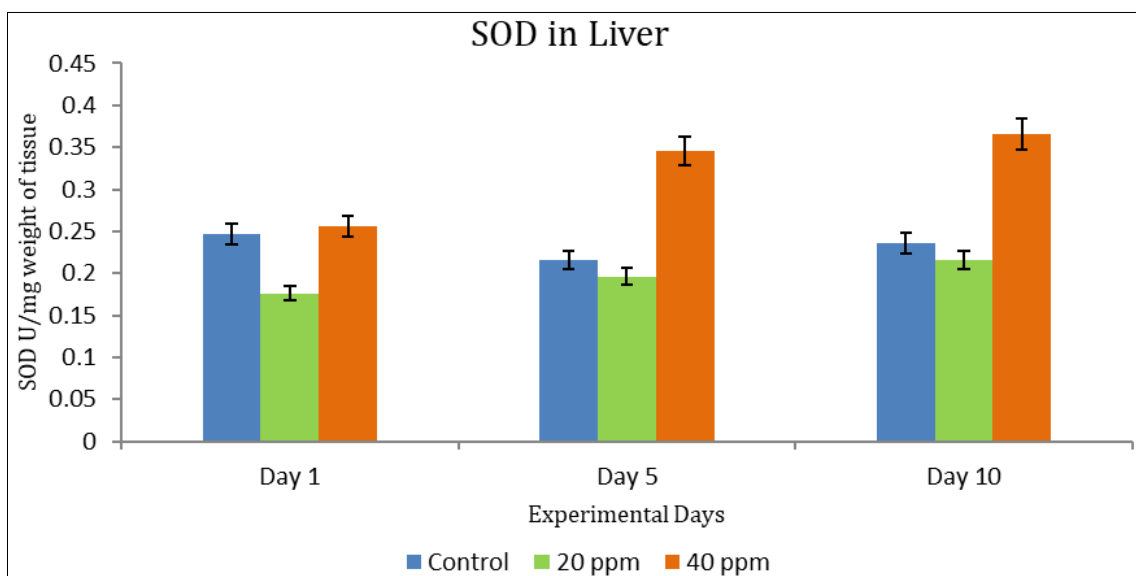
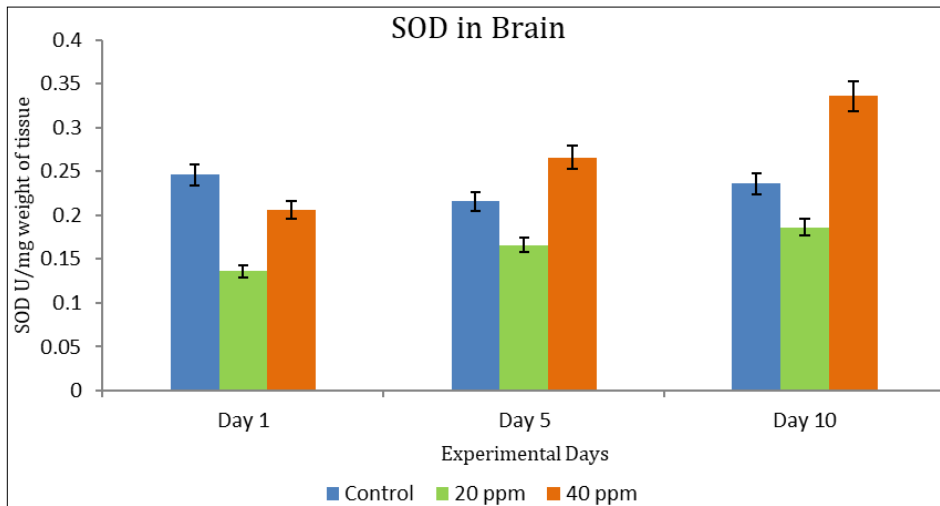


Fig 3: Comparative chart of SOD level in liver of *Channa punctatus*

**SOD in Brain**

Table 5: SOD level in Brain of *Channa punctatus*

Tissue	Day	Control	Experimental	
			20 ppm	40 ppm
Brain	1 <sup>st</sup>	0.25	0.14	0.21
	5 <sup>th</sup>	0.22	0.17	0.27
	10 <sup>th</sup>	0.24	0.19	0.34



**Fig 4:** Comparative chart of SOD level in Brain of *Channa punctatus*

## Discussion

### LPO

It is one of the main causes for the oxidative damage which is induced by metals [6].

It is used as biomarker of pollution [22].

Cd causes an enhancement in ROS formation, which leads to the peroxidation damage in liver and brain [26].

Liver shows an increased level of LPO on exposure to cadmium chloride due to an increased production of ROS.

Liver shows an increased level of LPO in 20ppm on 1<sup>st</sup>, 5<sup>th</sup> & 10<sup>th</sup> days 0.28<0.42<0.68 when compared with control (0.20).

Liver shows an increased level of LPO in 40ppm on 1<sup>st</sup>, 5<sup>th</sup> & 10<sup>th</sup> days 0.30<0.44<0.72 when compared with control (0.20).

Brain shows an increased level of LPO in 20ppm on 1<sup>st</sup>, 5<sup>th</sup> & 10<sup>th</sup> days 0.22<0.36<0.44 when compared with control (0.20).

Brain shows an increased level of LPO in 40ppm on 1<sup>st</sup>, 5<sup>th</sup> & 10<sup>th</sup> days 0.25<0.38<0.47 when compared with control (0.20).

Therefore, LPO was found to be low in control, further increased LPO was seen in experimental in liver followed by brain this is due to highest peroxidative damage.

### SOD

It is an antioxidant enzyme and also a group of metallo-enzymes, constitute a defense system against the natural or chemically induced production of ROS [19].

Liver shows decreased level of SOD in 20ppm on 1<sup>st</sup> day and increased in level was observed on 5<sup>th</sup> day & 10<sup>th</sup> day 0.18<0.20<0.22 when compared with control (0.22).

Liver shows decreased level of SOD in 40ppm on 1<sup>st</sup> day & increased in level was observed on 5<sup>th</sup> day & 10<sup>th</sup> day 0.26<0.35<0.47 when compared with control (0.22).

Brain shows decreased level of SOD in 20ppm on 1<sup>st</sup> day & increased in level was observed on 5<sup>th</sup> day & 10<sup>th</sup> day 0.14<0.17<0.19 when compared with control (0.22).

Brain shows decreased level of SOD in 40ppm on 1<sup>st</sup> day & increased in level was observed on 5<sup>th</sup> day & 10<sup>th</sup> day 0.21<0.27<0.34 when compared with control (0.22).

SOD: was found to be high in control and fluctuations was seen in experimental in liver followed by brain.

The result indicates that increased in LPO level and fluctuation in antioxidant defence system in fishes may be due to cadmium chloride toxicity.

Therefore the effect of Cadmium chloride on LPO & SOD depends on the Cd chloride concentrations (20ppm & 40ppm) and also with different time intervals (1<sup>st</sup> day, 5<sup>th</sup> day, & 10<sup>th</sup> day).

### Conclusion

Cadmium has an adverse effect on the respiratory, reproductive and haematological system in many fishes [24]. The present study revealed that Cadmium chloride toxicity is responsible for the oxidative stress in liver and brain tissues of *Channa punctatus* as shown by increase in LPO and in response, the antioxidant (SOD) defense mechanism were induced.

Oxidative stress may cause aging processes and also diseases like Parkinson's, Cancer and Alzheimer's.

Therefore it is advised for the patients who are suffering from diseases to take foods rich in Antioxidants to overcome with oxidative damage.

### Abbreviations

**DETPA:** Diethylenetriaminepentaacetic acid

**DNA:** Deoxyribonucleic acid

**EDTA:** Ethylenediaminetetraacetic acid

**HCL:** Hydrochloric acid

**LPO:** Lipid peroxidation

**ROS:** Reactive oxygen species

**SOD:** Superoxide dismutase

**TBA:** Thiobarbituric acid

**TCA:** Trichloroacetic acid

### Acknowledgement

The satisfaction and euphoria that accompany the successful completion of this task would be incomplete without mentioning the people involved, whose constant guidance and encouragement made it possible. It is my pleasure to acknowledgement the help of all those individuals who have extended their co-operation in various ways during my project work.

I express my profound gratitude to my guide Dr. C. Manjusha, Department of Zoology for her guidance, constructive suggestions and encouragement in completing my project work.

I avail this opportunity to express my gratitude to my Principal Dr. Shobhana Deshpande, PG Director Subba Rao (Department of Computer Science), Dr. S. Srivani (Head of Department Zoology), Dr. Manjula M. Reddy (Department of Zoology (PG)), Dr. D. Suneetha Devi (Department of Zoology (PG)), Sarojini Naidu Vanita Maha Vidyalaya for providing their invaluable guidance, encouragement and suggestions throughout the course of the project.

I would like to thank my parents and also my friends for helping and supporting me throughout the course of my project.

## References

1. Almroth BC, Sturve J, Berglund A, Forlin L. Oxidative damage in eelpout (*Zoarces viviparus*), measured as protein carbonyls and TBARS, as biomarkers. *Aquat Toxicol*,2005;73:171–180.
2. AMAP. Assessment report: Arctic pollution issues Arctic monitoring and assessment programme, 1998.
3. Cooper CE, Vollaard NBJ, Choueiri T, *et al.* Exercise, free radicals and oxidative stress. *Biochem Soc Trans*,2002;30(2):280-5.
4. Dekkers JC, van Doornen LJ, Kemper HC. The role of antioxidant vitamins and enzymes in the prevention of exercise-induced muscle damage. *Sports Med*,1996;21(3):213-38.
5. Draper HH, Squires EJ, Mahmooch H, Wu J, Agarwal S, Handley M. A comparative evaluation of thiobarbituric acid methods for the determination of malondialdehyde in biological materials. *Free Radic Biol Med*,1993;15:353–363.
6. Ercal N, Gurer Orhan H, Aykin Burns N. Toxic metals and oxidative stress Part I: Mechanisms involved in metal induced oxidative damage. *Current Topics in Medicinal Chemistry*,2001;1:529–539.
7. IARC (International Agency for Research on Cancer). Monographs on the Evaluation of the Carcinogenic Risks to Humans Beryllium, Cadmium, Mercury and Exposures in the Glass Manufacturing Industry; IARC Scientific Publications: Lyon, France, 1993, 119–238.
8. Kohen R, Nyska A. Oxidation of biological systems: oxidative stress phenomena, antioxidants, redox reactions, and methods for their quantification. *Toxicol Pathol*,2002;30:620–650.
9. Kumar R, Nagpure NS, Kushwaha B, Srivastava SK, Lakra WS. Investigation of the genotoxicity of Malathion to freshwater teleost fish *Channa punctatus* (Bloch) using the micronucleus test and comet assay. *Arch. Environ. Contam. Toxicol*,2010;58:123–130.
10. Livingstone DR. Contaminant reactive oxygen species production and oxidative damage in aquatic organisms. *Mar Pollut Bull*,2001;42:656–666.
11. Livingstone DR. Oxidative stress in aquatic organism in relation to pollution and aquaculture. *Revue de Medecine Veterinaire*,2003;154:427–430.
12. Luoma SN, Rainbow PS. Sources and cycles of trace metals. In: *Metal Contamination in Aquatic Environments: Science and Lateral Management*. Cambridge University Press, Cambridge, 2008a, 47–66.
13. Lushchak V, Lushchak LP, Mota AA, Hermes Lima M. Oxidative stress and antioxidant defences in goldfish *Carassius auratus* during anoxia and reoxygenation. *Am J Physiol Regul Integr Comp Physiol*,2001;280:100–107
14. Matovic V, Buha A, Đukić Cosić D, Bulat Z. Insight into the oxidative stress induced by lead and /or cadmium in blood, liver and kidneys. *Food Chem. Toxicol*,2015;78:130–140. [CrossRef]
15. Nishida Y. The chemical process of oxidative stress by copper (II) and iron (III) ions in several neurodegenerative disorders. *Monatshefte fur Chemie*,2011;142:375–384.
16. Nishijo M, Satarug S, Honda R, Tsuritani I, Aoshima K. The gender differences in health effects of environmental cadmium exposure and potential mechanisms. *Mol. Cell. Biochem*,2004;255:87–92. [CrossRef]
17. Pizzino G, Bitto A, Interdonato M, Galfo F, Irrera N, Mecchio A, *et al.* Oxidative stress and DNA repair and detoxification gene expression in adolescents exposed to heavy metals living in the Milazzo-Valle del Mela area (Sicily, Italy). *Redox Biol*,2014;2:686–693. [CrossRef] [PubMed]
18. Powers SK, Lennon SL. Analysis of cellular responses to free radicals: focus on exercise and skeletal muscle. *Proc Nutr Soc*,2000;58:1025-33.
19. Roche H, Boge G. Fish blood parameters as a potential tool for identification of stress caused by environmental factors and chemical intoxication. *Mar Environ Res*,1996;41:27–43.
20. Satarug S, Swaddiwudhipong W, Ruangyuttikarn W, Nishio M, Ruiz P. Modeling cadmium exposures in low- and high-exposure areas in Thailand. *Environ. Health Perspect*,2013;121:531–536. [CrossRef]
21. Satarug S, Vesey DA, Gobe GC. Health risk assessment of dietary cadmium intake: Do current guidelines indicate how much is safe? *Environ. Health Perspect*,2017;125:284–288. [CrossRef]
22. Sayeed I, Parvez S, Pandey S, Bin Hafeez B, Haque R, Raisuddin S. Oxidative stress biomarkers of exposure to deltamethrin in freshwater fish, *Channa punctatus* Bloch. *Ecotoxicol Environ Saf*,2003;56:295–301.
23. Sies H. Strategies of antioxidant defence. *Eur J Biochem*,1993;215:213–219.
24. Soares SS, Aureliano M, Joaquim N, Coucelo JM. Cadmium and vanadate oligomers effects on methaemoglobin reductase activity from Lusitanian toadfish: *in vivo* and *in vitro* studies. *J Inorg Biochem*,2003;94:285–290.
25. Soares SS, Martins H, Gutierrez Merino C, Aureliano M. Vanadium and cadmium *in vivo* effects in teleost cardiac muscle: metal accumulation and oxidative stress markers. *Comp Biochem Physiol C*,2008;147:168–178.
26. Stohs SJ, Hagchi D, Hassoun E, Bagshi M. Oxidative mechanisms in the toxicity of chromium and cadmium ions. *J Environ Pathol Toxicol Oncol*,2000;19:201–213.
27. Thompson J, Bannigan J. Cadmium: Toxic effects on the reproductive system and the embryo. *Reproduct. Toxicol*,2008;25:304–315. [CrossRef] [PubMed]
28. Wang J, Zhu H, Liu X, Liu Z. Oxidative stress and Ca<sup>2+</sup> signals involved on cadmium-induced apoptosis in rat hepatocyte. *Biol. Trace Elem. Res*,2014;161:180–189. [CrossRef]
29. WHO World Health Organization. *Environmental Health Criteria 134: Cadmium*; World Health Organization: Geneva, Switzerland, 1992.